

dynamic BIOSENSORS

# **PURIFICATION COLUMN**

**Description and handling instructions** 

Dynamic Biosensors GmbH & Inc. PF-CC-1 v5.1





## **Product Description**

Order Number: PF-CC-1

Table 1. Contents and Storage Information

Column Specifications		Media Specifications	
Column Volume	1 mL	Matrix	Hydrophilic porous polymer beads
Column material	Polypropylene	Particle size	30 μm
Column size length x I.D.	26 x 7.0 mm	Functional group	-R-N <sup>+</sup> (CH <sub>3</sub> ) <sub>3</sub>
Recommended flow rate	1 mL/min	pH range	2 - 12
Maximum flow rate	4 mL/min	Temp. range	4 - 60 °C
Max. pressure	0.3 MPa	Shipping solvent	20 % EtOH aqueous solution

### **Handling**

#### Storage and Lifetime

Store in 20 % ethanol at 4 - 35 °C.

For longer lifetime, flush weekly the column with water, then with 20 % EtOH solution. Make sure to close the end tightly to avoid drying out.

#### **Equilibration and Elution**

Protein-DNA conjugates are electrostatically bound to the matrix of the column with **pro**FIRE® Buffer A as a first mobile phase, then eluted with a salt-concentration gradient method (**pro**FIRE® Buffer B).

Water-soluble organic solvent (maximum of 30 %) can be added in the mobile phase. Before adding such solvent, make sure that the salt will not precipitate.

# **Cleaning / Troubleshooting**

If the column is showing a change in retention time of the free DNA peak or alterations in peak shape and/or pressure increase, follow the washing steps described below.

- 1. Place buffer tubing A and B in a bottle with water (filtered) and start a "Clean & Sleep" run via Mobile Control software. Leave the column in.
- 2. Place the buffer **tubing B** in a bottle with **0.1 M to 0.5 M NaOH** (filtered) and leave the buffer tubing A in water (filtered).
- 3. **Reverse the column** and connect the column outlet to the inlet-tubing coming from the pump. Do not connect the other outlet of the column to the tubing going to the flow cell.
- 4. Place the opened outlet of the column in a clean bottle to collect the washing solution.

**NOTE** It should be avoided contaminating the system with the washing solution.

- 1. Tap the "System Overview" symbol, then tap on "Sample injection" in Mobile control software.
  - a. Check if the **fraction valve port** is set to **position 13** (waste position)
  - b. Check if the **injection valve port** is set to **position 1** (bypass position)
  - c. Select the pump on the left bottom side
  - d. Set the pump parameters for channel B: Select channel B (100%), adjust the flow to 1 mL/min, tap "Apply" and



"Run".

- 2. Collect the fluid eluting from the column and leave the flow for 10 15 minutes.
- 3. Set the pump parameters for channel A: Select channel A (100%), adjust the flow to 1 mL/min, tap "Apply" and "Run".
- 4. Collect the fluid eluting from the column and leave the flow for 15 minutes.
- 5. Stop the flow and **reverse the column** in its original intended position with both PEEK-tubings connected to the system.
- 6. Put both buffer tubing A and B in a bottle with water (filtered) and start two "Clean & Sleep" runs, one after each other via Mobile Control software.
- 7. Place the tubing A in (filtered and degassed) **pro**FIRE® Buffer A [1] and the tubing B in (filtered and degassed) **pro**FIRE® Buffer B [2] and start a buffer run.

  Type in the **DNA length 1, inject 160 μL** of **pro**FIRE® Buffer A and press **"Start"**.
- 8. Start a DNA only run with a known DNA oligo you can refer to.



# **Useful Order Numbers**

Table 2. Order Numbers

Product Name	Amount	Order No
10x Buffer A [1]	50 mL (yielding 500 mL)	PF-BU-A-10
5x Buffer B <sup>[2]</sup>	50 mL (yielding 250 mL)	PF-BU-B-5
proFIRE® Amine Coupling Kit 1 for proteins (>5 kDa);	5 conjugations	PF-NH2-1
proFIRE® Thiol Coupling Kit 1 for proteins (>5 kDa);	5 conjugations	PF-SH-1



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<sup>[1]</sup> Buffer A: 50 mM Na<sub>2</sub>HPO<sub>4</sub>/NaH<sub>2</sub>PO<sub>4</sub>, 150 mM NaCl, pH 7.2