

Theory Note

Association rate of two equimolar reactants in solution

Keywords: Association kinetics | solution phase | second order reaction



Both reactants are present at equal concentrations at $t=0$

$$[A]_0 = [B]_0 = c_0 \quad (2)$$

Second order reaction rate law

$$\frac{dc}{dt} = -k_{on}c^2 \quad (3)$$

Integrated second order rate law

$$\frac{1}{c_t} = \frac{1}{c_0} + k_{on}t \quad (4a)$$

$c_t=c(t)$ = concentration of unbound reactant

$$c_t = \frac{c_0}{1 + c_0k_{on}t} \quad (4b)$$

Units of rate constant k_{on}

$$M^{-1}s^{-1}$$

For the measurement of the association reaction in the solution phase (e.g. stopped flow in a cuvette) it is assumed that a signal S (e.g. fluorescence, absorption) is monitored over time. At time $t=0$, the reactants are mixed (but could not react yet) and the signal is S_0 . At time $t=\infty$, an equilibrium has established (all reactants are bound) and the signal is S_∞ . The change in signal S over time (S_t) is described by:

Signal at time t

$$S_t = \underbrace{S_0}_{\text{start value}} + \underbrace{(S_\infty - S_0)}_{\text{amplitude of signal change upon binding}} \cdot \underbrace{\left(\frac{c_0 - c_t}{c_0}\right)}_{\text{fraction bound}} \quad (5)$$

$$S_t = S_0 + (S_\infty - S_0) \cdot \left(1 - \frac{c_t}{c_0}\right)$$

(4b) in (5b)

$$S_t = S_0 + (S_\infty - S_0) \cdot \left(1 - \frac{1}{1 + c_0k_{on}t}\right) \quad (6a)$$

Signal as a function of time

$$S_t = S_\infty - \frac{S_\infty - S_0}{1 + c_0k_{on}t} \quad (6b)$$

alternative representation of
(6b)

$$S_t = S_0 + \frac{(S_\infty - S_0)c_0 k_{on} t}{1 + c_0 k_{on} t} \quad (6c)$$

Linear plot to determine k_{on}

$$\frac{1}{S_\infty - S_t} = \frac{1}{S_\infty - S_0} + \frac{c_0 k_{on}}{S_\infty - S_0} \cdot \frac{t}{x} \quad (7)$$

Example

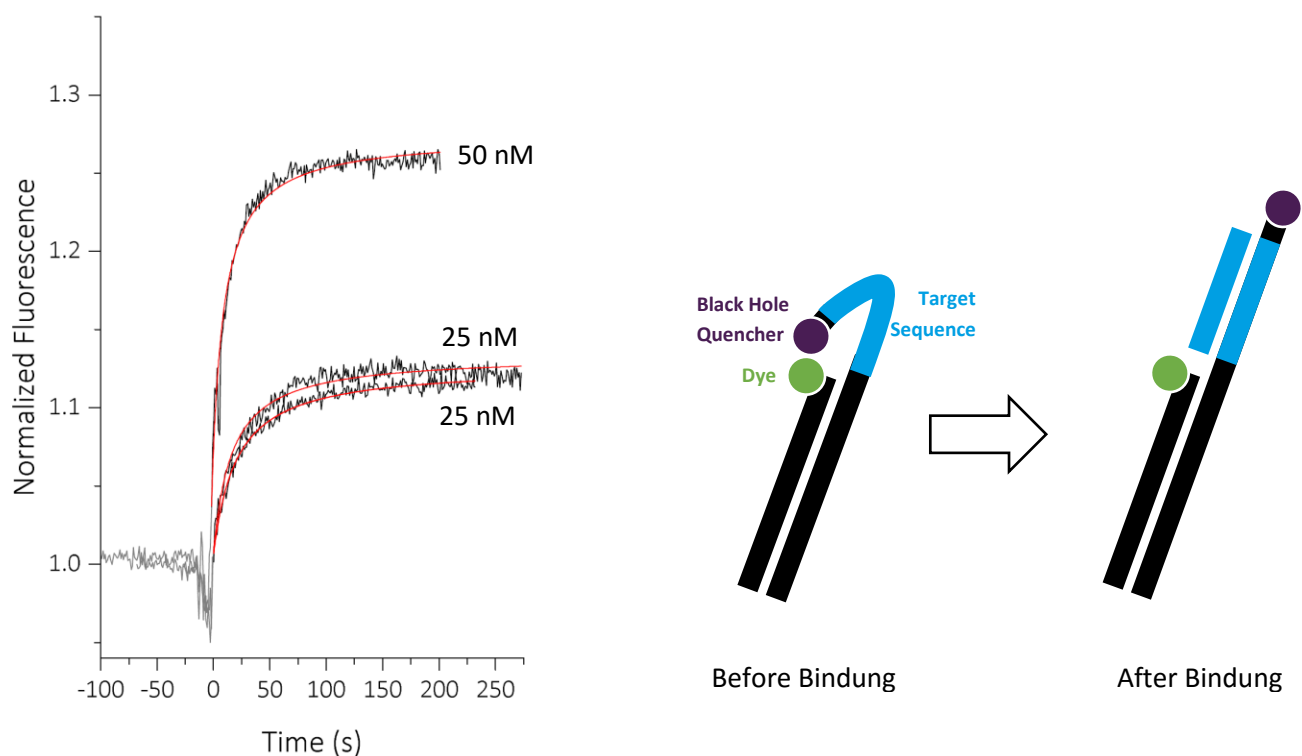


Figure 1 | Formation of a 10 bp DNA duplex in the solution phase.

Reactant 1: 17 base single stranded DNA, labelled with a green fluorophore at one end and a black hole quencher at the opposing end.
 Reactant 2: 10 base single strand, complementary to the overhang of reactant 1 (unlabeled). At $t=0$, reactant 2 was injected into a cuvette containing reactant 1 under intense stirring; final concentrations of both reactants were 25 nM (equimolar). The fluorescence increase (measured with a Shimadzu RF5301 spectrofluorophotometer) indicates the formation of the 10 bp duplex in real-time, because the quenching (FRET) efficiency decreases upon formation of duplex DNA, because it constitutes an effective rod-like spacer between the fluorophore and the quencher.

The solid red line is a non-linear least-squares fit with equation 6, yielding the association rate constant $k_{on} = 2.0 \cdot 10^6 \text{ M}^{-1} \text{ s}^{-1}$.

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